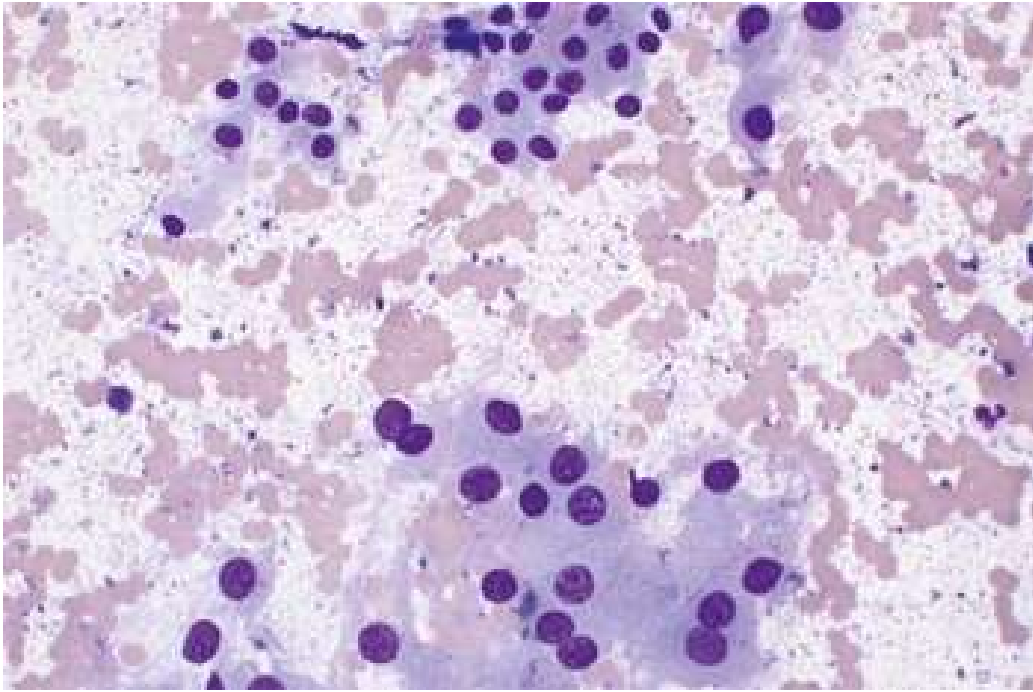


# Nordic immunohistochemical Quality Control



EQALM Symposium 2025 – 16.-18. October – Vienna

Rasmus Røge, MD, PhD  
Clinical Associate Professor, Aalborg University Hospital  
NordIQC scheme organizer, Senior registrar,  
Department of Pathology  
Aalborg University Hospital, Denmark



# Cytology

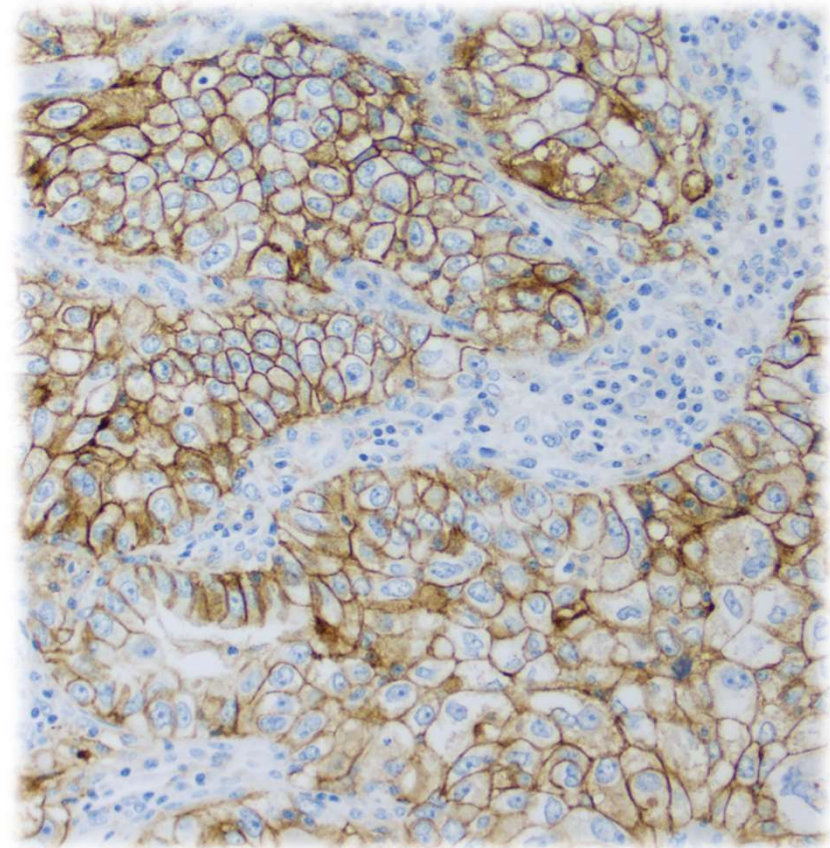
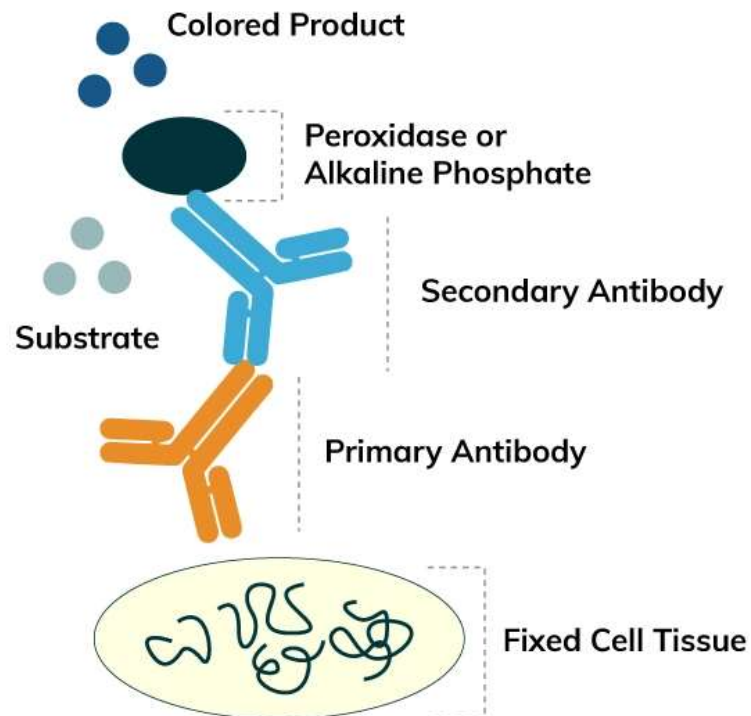
MGG



# Histology

HE

# Immunohistochemistry (IHC)

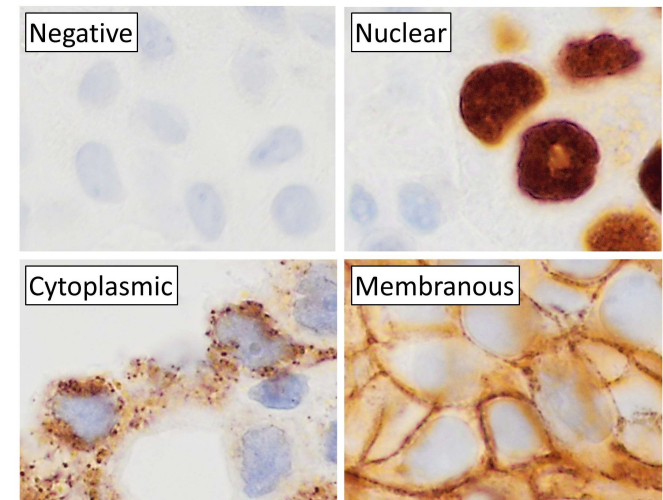
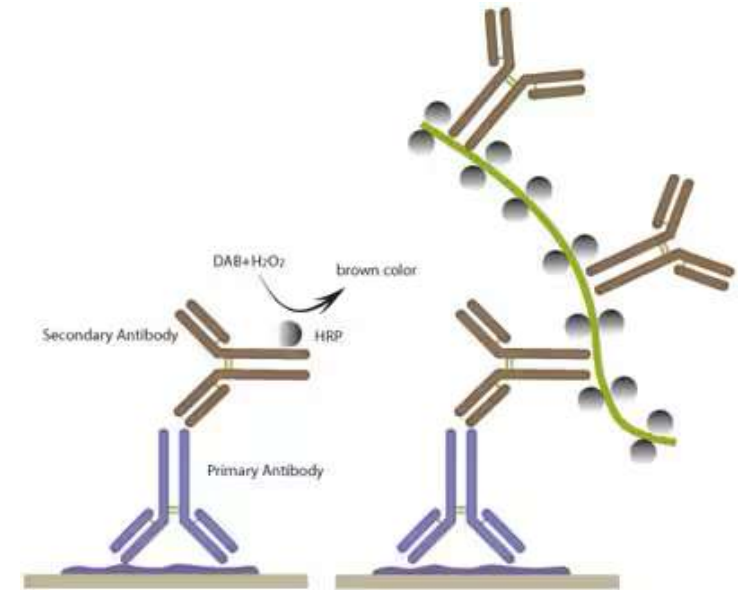


# Immunohistochemistry (IHC)

“Spatial proteomics”

Most abundant “stain”  
after HE

100-300 different  
epitopes in most labs

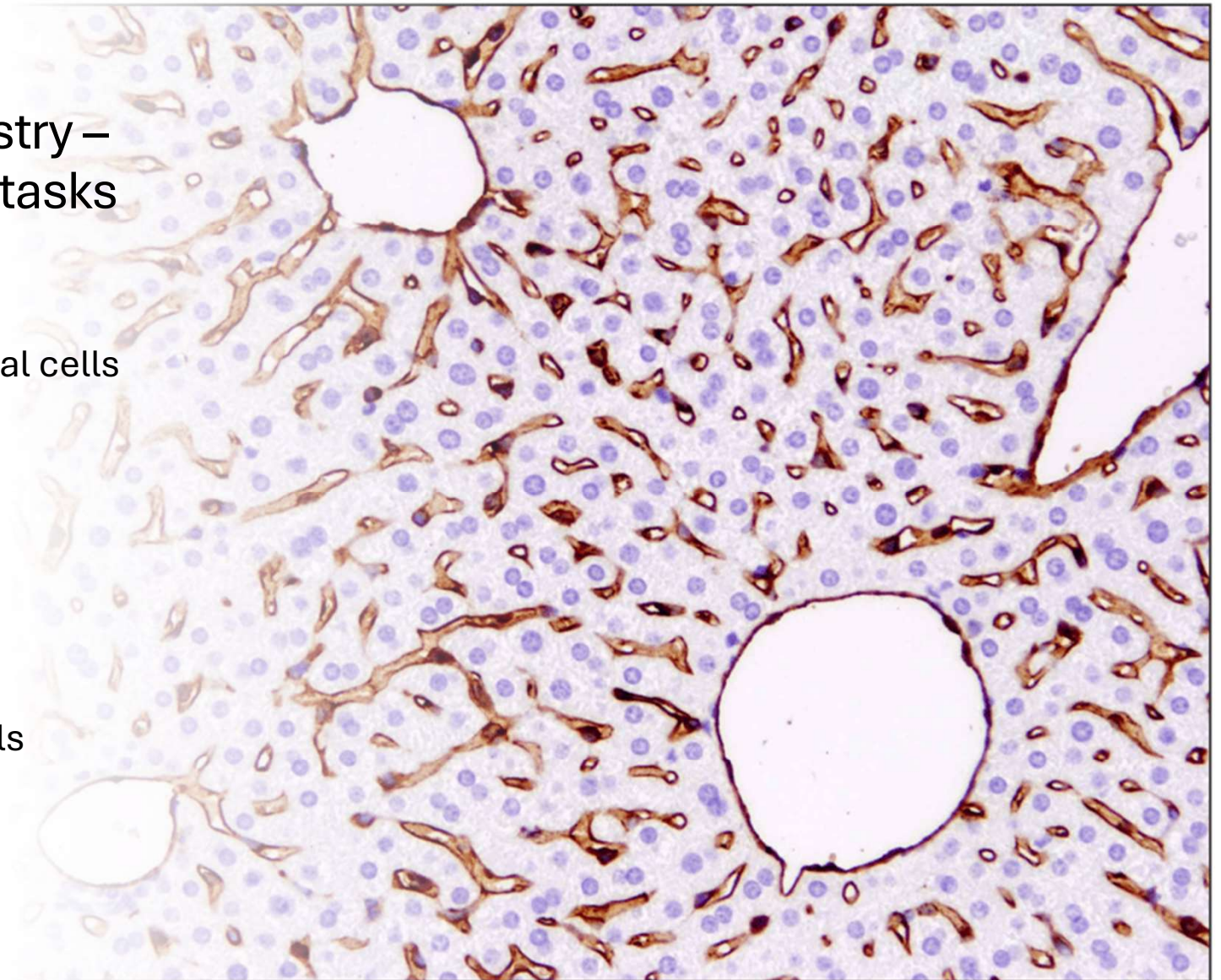


# Usages of IHC in clinical pathology

- Detection of specific proteins in patient materials that will help to confirm or disprove a diagnosis
- Identify origin of unknown primary or metastatic cancer
- Subclassify cancers (prognostic)
- Predictive biomarkers

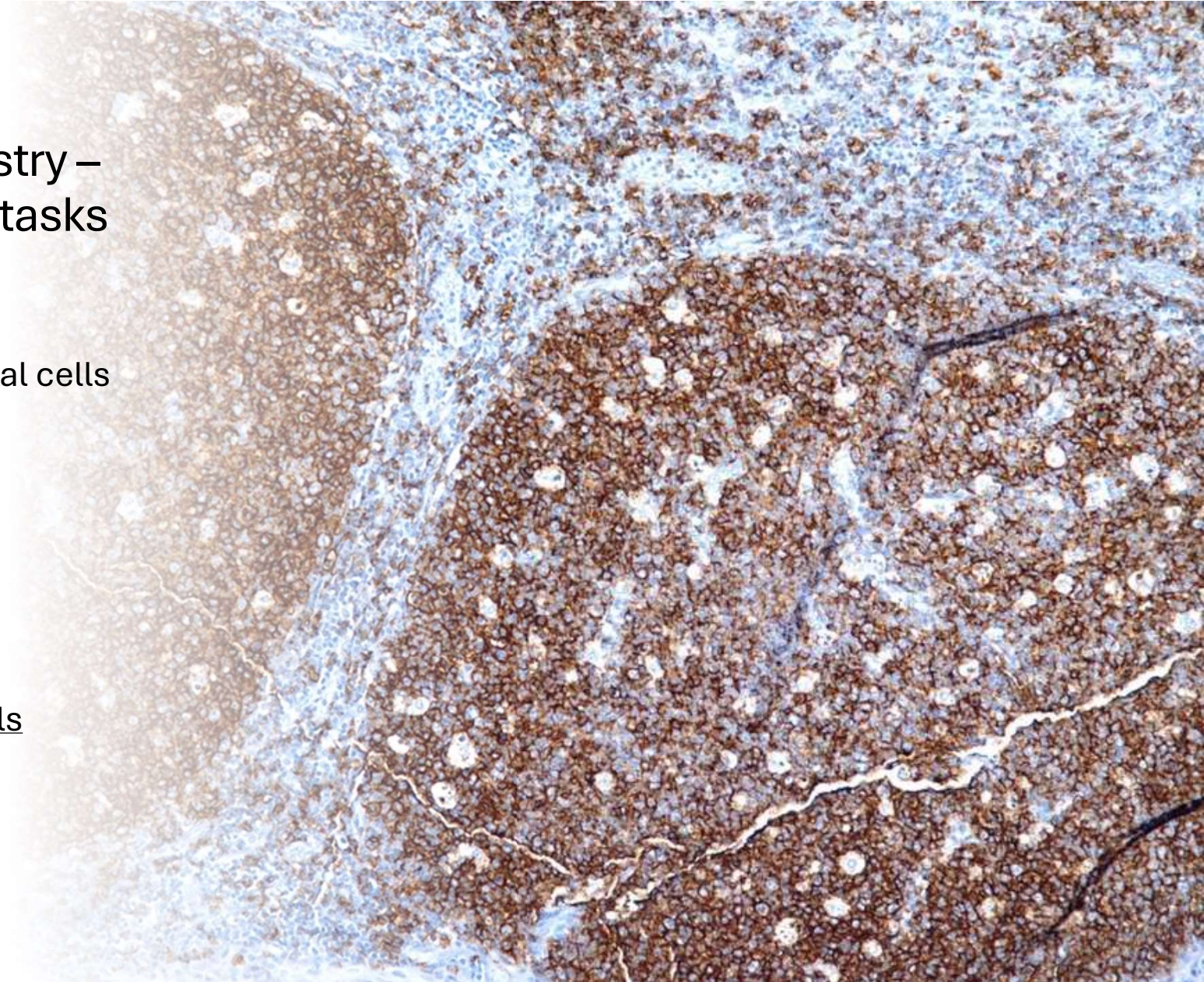
## Immunohistochemistry – examples of simple tasks

- Pan cytokeratin: Epithelial cells
- CD31: vessels
- Ki67: proliferation
- CD3/CD20: T- and B-cells
- CD34: Stem cells

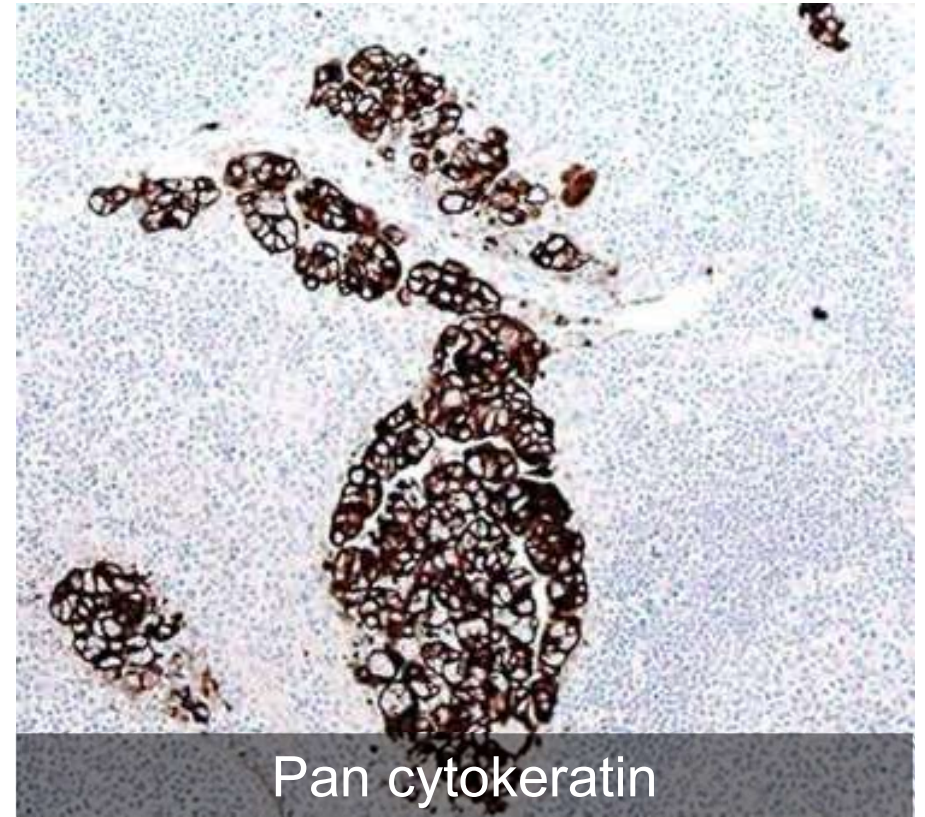
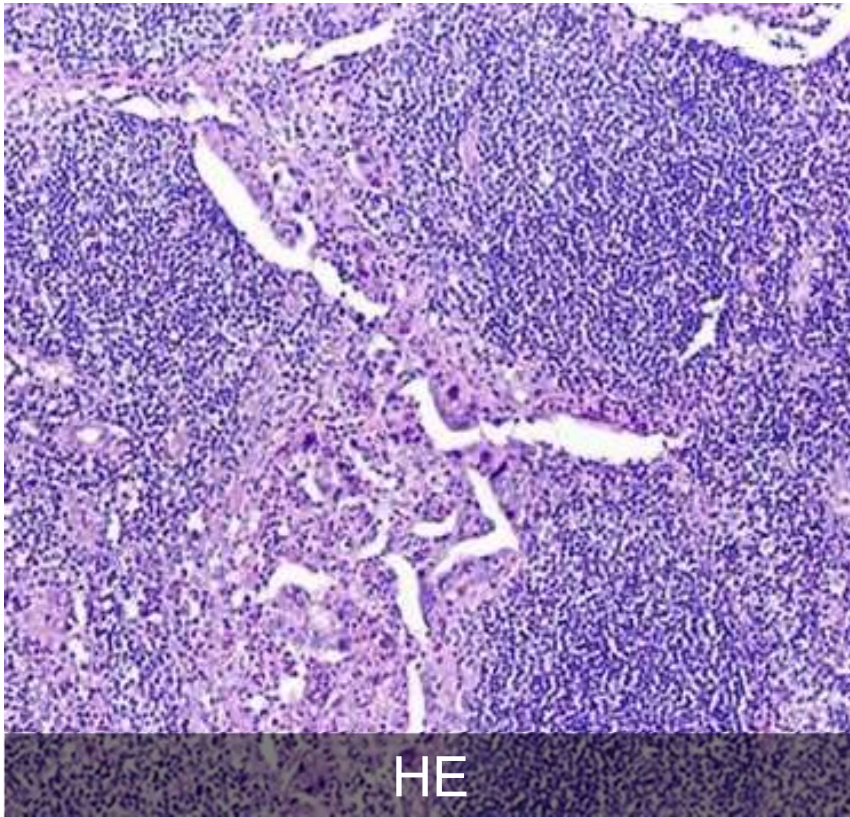


## Immunohistochemistry – examples of simple tasks

- Pan cytokeratin: Epithelial cells
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- CD3/CD20: T- and B-cells
- CD34: Stem cells



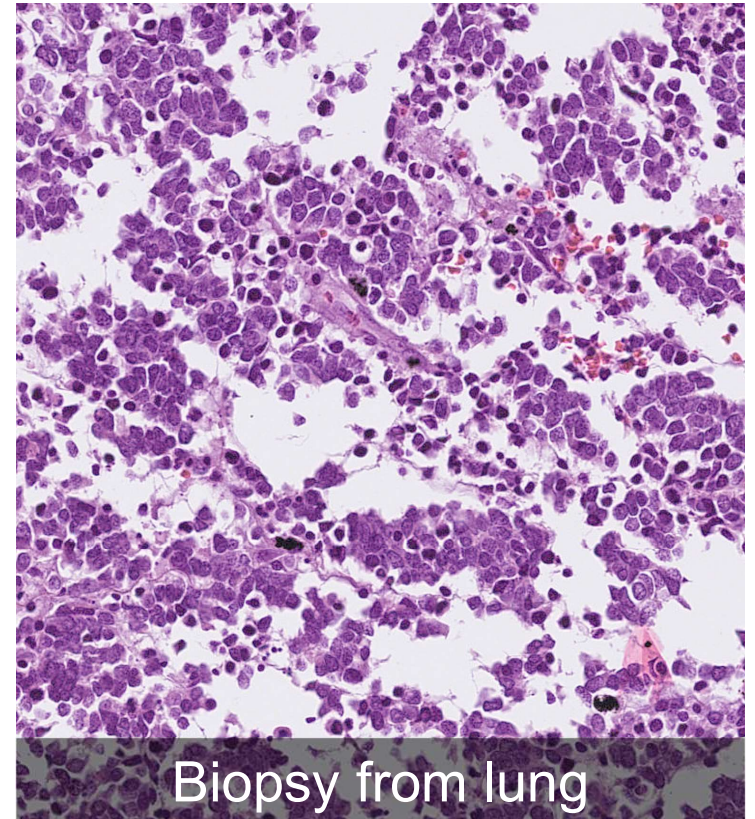
# Pan cytokeratin - micrometasis





# Case

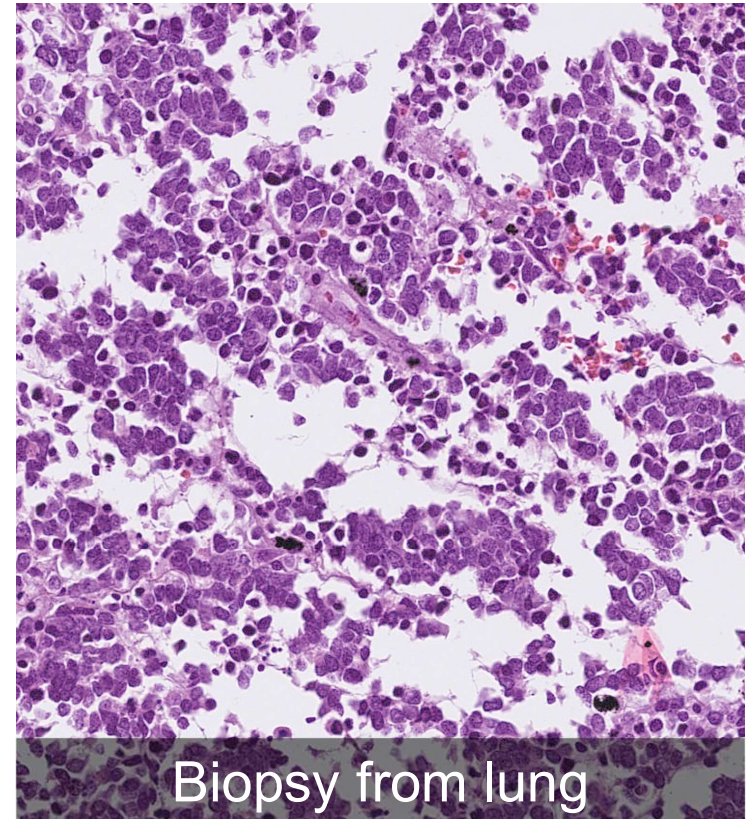
- 67 y.o. female
- Heavy smoker
- Several tumours in both lungs
- Pleural plaques and exposed to asbestos
- Previous ovarian serous carcinoma



# Case

- Carcinoma
- Lung cancer ?
  - Adeno? Squamous? Small cell (neuroendocrine)?
- Relapse ovarian serous carcinoma ?
- Mesothelioma ?

CD45	Pan cytokeratin	S100	Vimentin
Negative	Positive	Negative	Negative



# Selected panels

Lung:  
TTF  
Napsin

Neuroendocrine:  
SYP  
CGA  
INSM1

Female genitals:  
PAX8  
WT1  
ER

Mesothelioma:  
Calretinin  
Podoplanin

## Lung

TTF	Positive
Napsin	Negative

## Neuroendocrine

SYP	Positive
CGA	Positive
INSM1	Positive

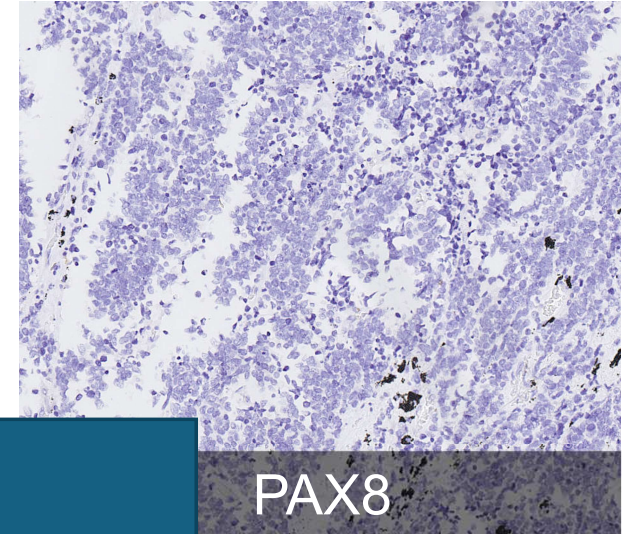
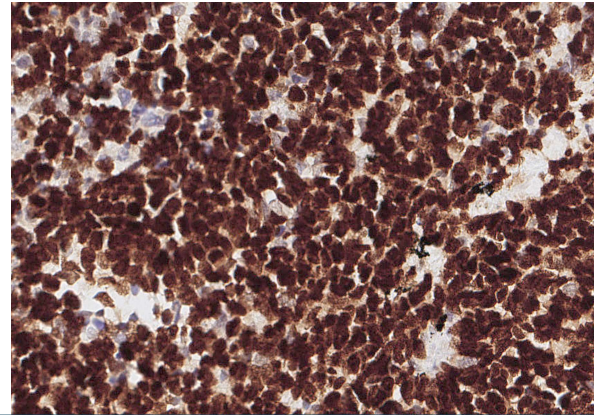
## Female genitals

PAX8	Negative
WT1	Negative
ER	Negative

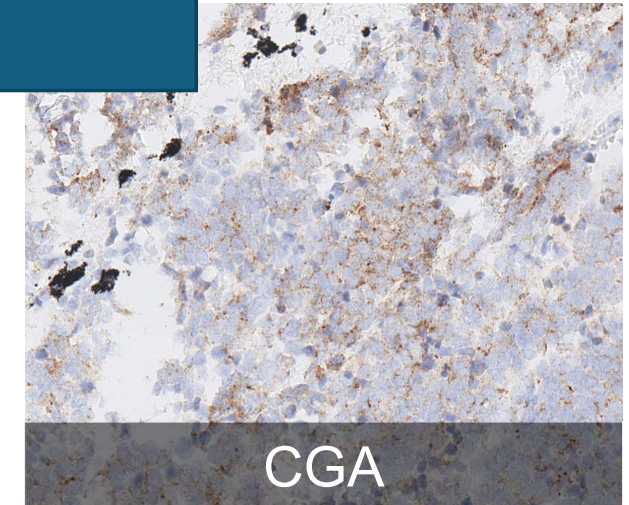
## Mesothelioma

Calretinin	Negative
Podoplanin	Negative

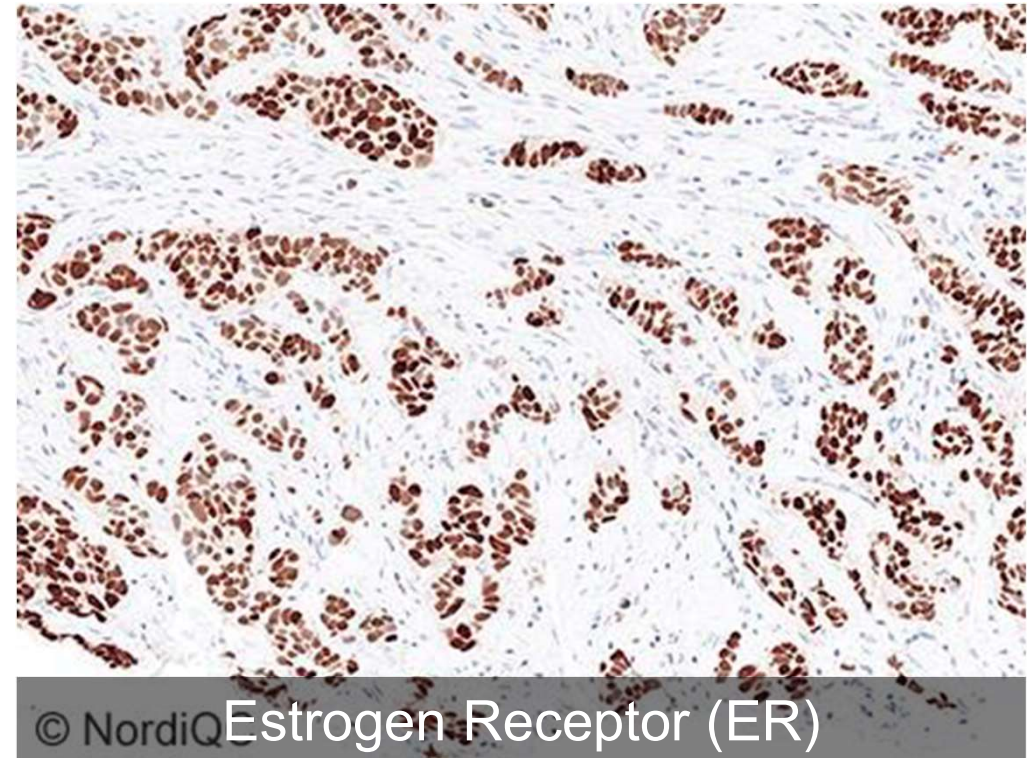
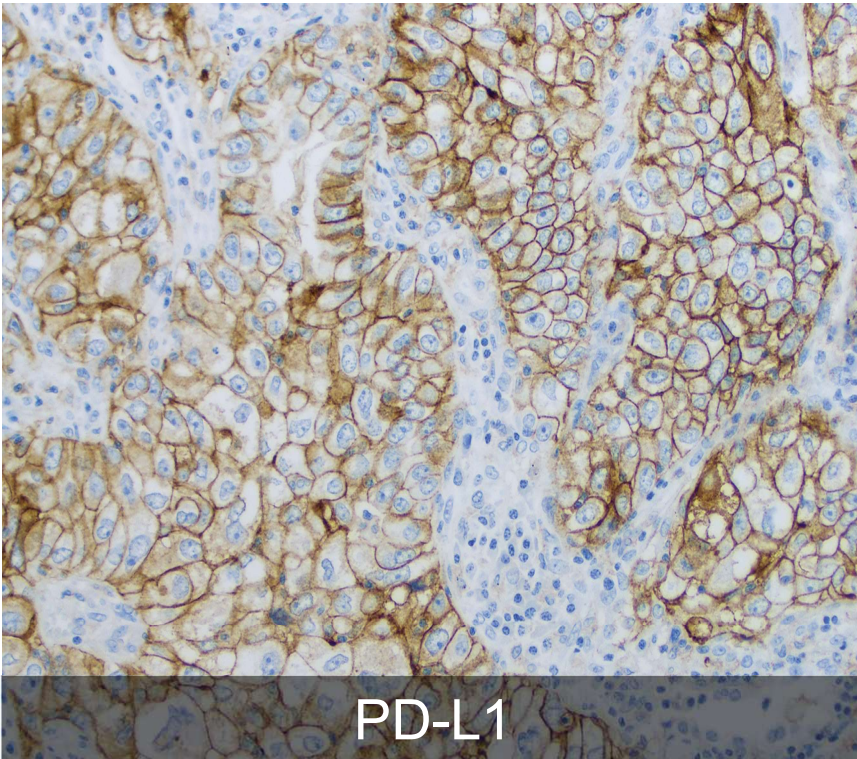
# Case



Small cell lung carcinoma  
(neuroendocrine)



# Predictive biomarkers



# FDA approved IHC assays

Biomarker	Indication	Drug	Test	Product
HER2	Breast cancer, Gastric ad carc.	Trastuzumab, Pertuzumab,...	IHC	HercepTest™, PATHWAY®
ALK	NSCLC	Crizotinib, Ceritinib, ...	IHC	VENTANA ALK D5F3 assay
CD117	Gastrointestinal stromal tum.	Gleevec	IHC	Dako c-KIT pharmDx
MMR	Endometrial carcinoma	Dostarlimab-gxly	IHC	VENTANA MMR RxDx panel
PD-L1	NSCLC, Urothelial carc. HNSCC, TNBC, ESCC, Cervical cancer.	Pembrolizumab	IHC	Dako 22C3 IHC pharmDX
PD-L1	NSCLC, (Melanoma optional)	Nivolumab	IHC	Dako 28-8 IHC pharmDX
PD-L1	NSCLC, Urothelial carc.	Atezolizumab	IHC	VENTANA PD-L1 SP142
PD-L1	Urothelial carc., (NSCLC)	Durvalumab	IHC	VENTANA PD-L1 SP263

“Immunohistochemistry is technically complex, and no aspect of this complexity can be ignored, from the moment of collecting the specimen to issuance of the final report “  
Taylor CR. Arch Pathol Lab Med 2000; 124:945

### Pre-Analytical

Ischemia  
Fixation process  
(Decalcification)  
Tissue processing  
Paraffin embedding  
Sectioning  
Storage



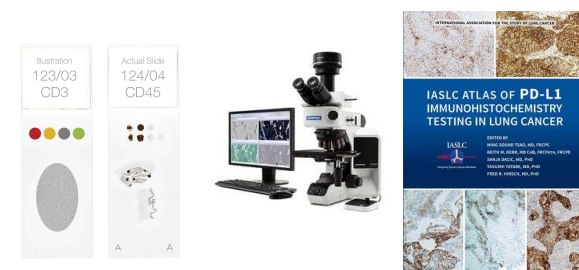
### Analytical

IHC platform  
Epitope retrieval  
Primary antibody  
Detection system  
Chromogen  
Counterstaining  
Mounting



### Post-Analytical

Usage of controls  
Positive controls  
Negative controls  
“Critical controls”  
Scoring / read-out  
Interpretation  
Reporting

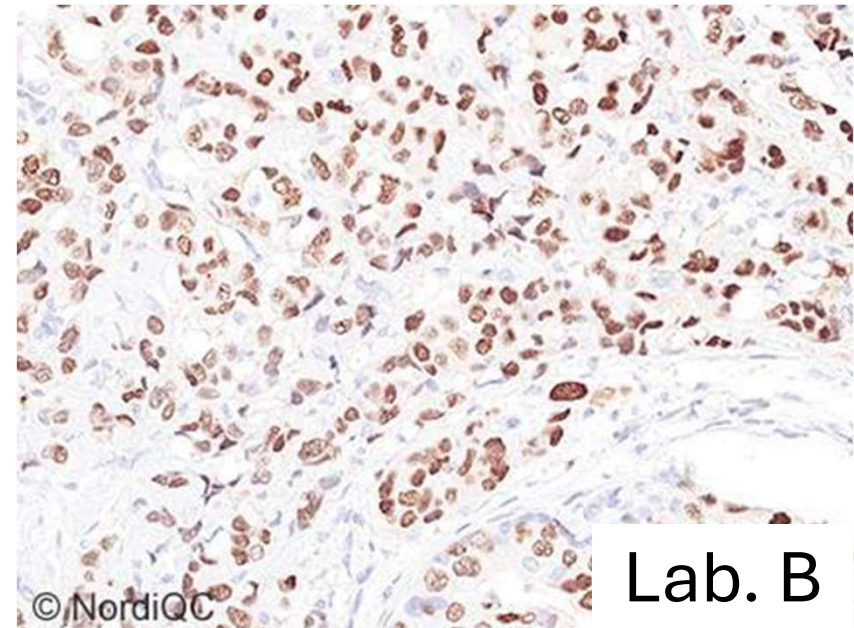
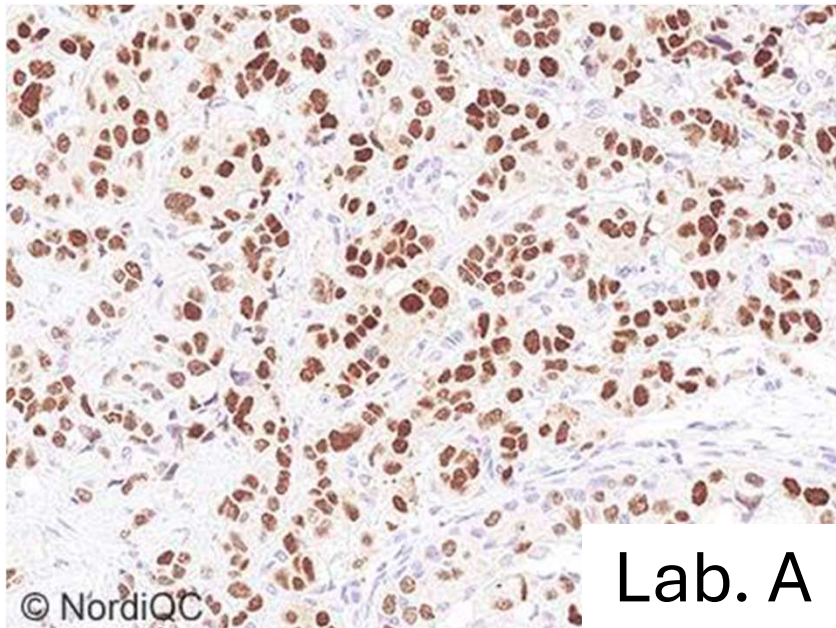


# Nordic immunohistochemical Quality Control (NordiQC)

- External Quality Assessment scheme focusing on the analytical phase of IHC
- Assessing the IHC assay quality in international pathology labs
  - Based on “standard” processed circulated tissues
- Identifying optimal and insufficient results
  - Correlated to antibodies, protocols and stainer platforms
- Giving directions for improvement
  - Individually tailored recommendations

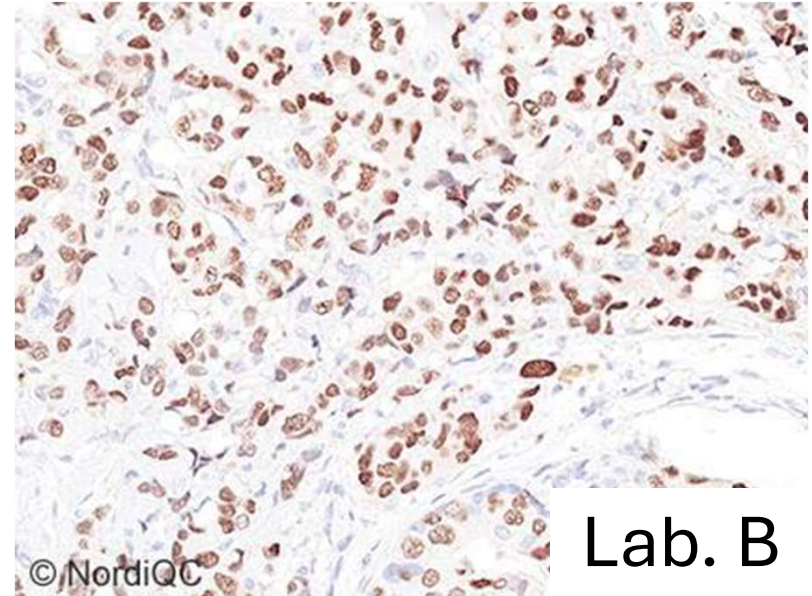
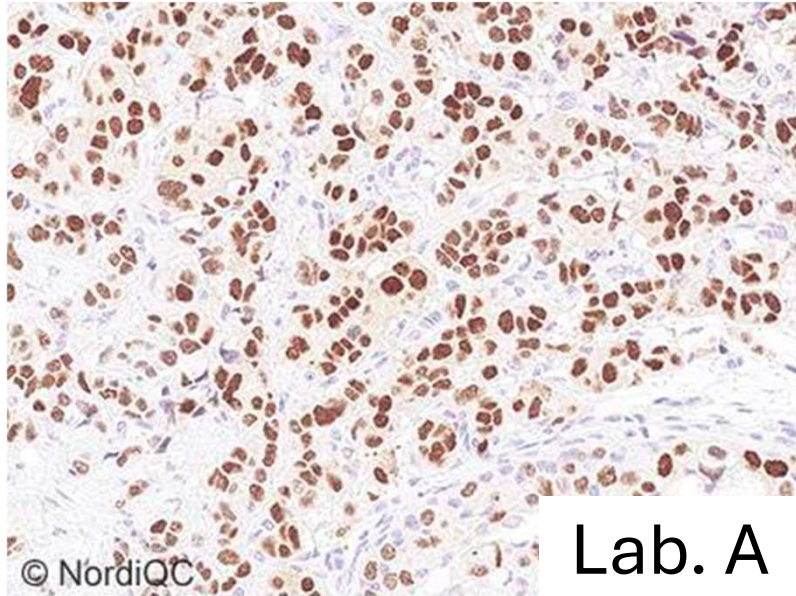


# Serial sections (breast carcinoma) stained for Estrogen Receptor (ER)

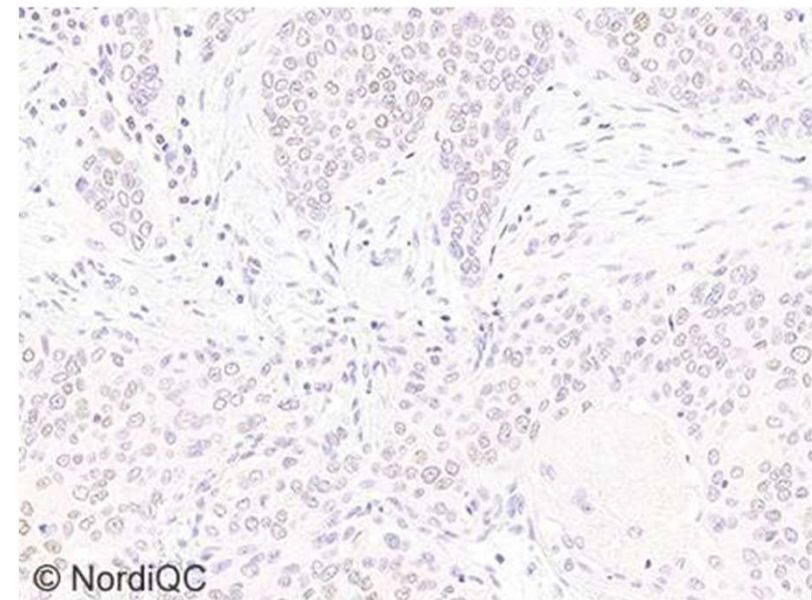
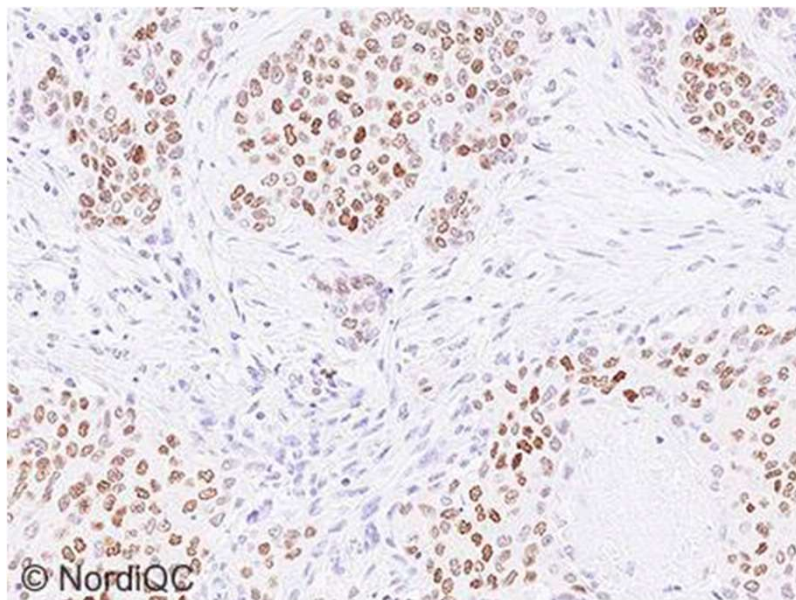


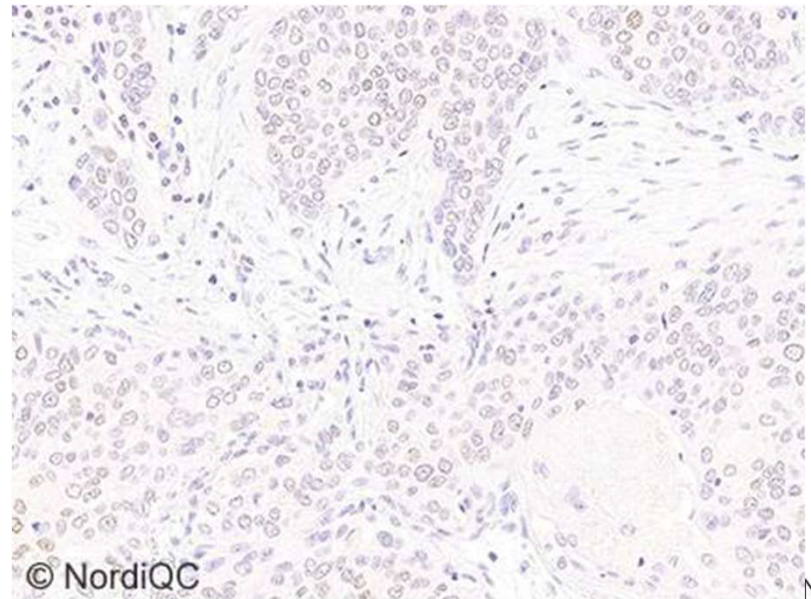
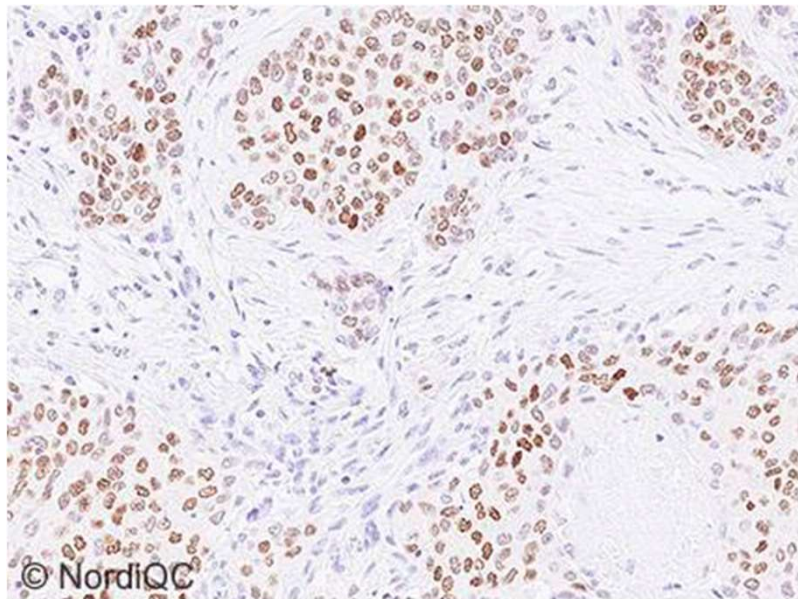
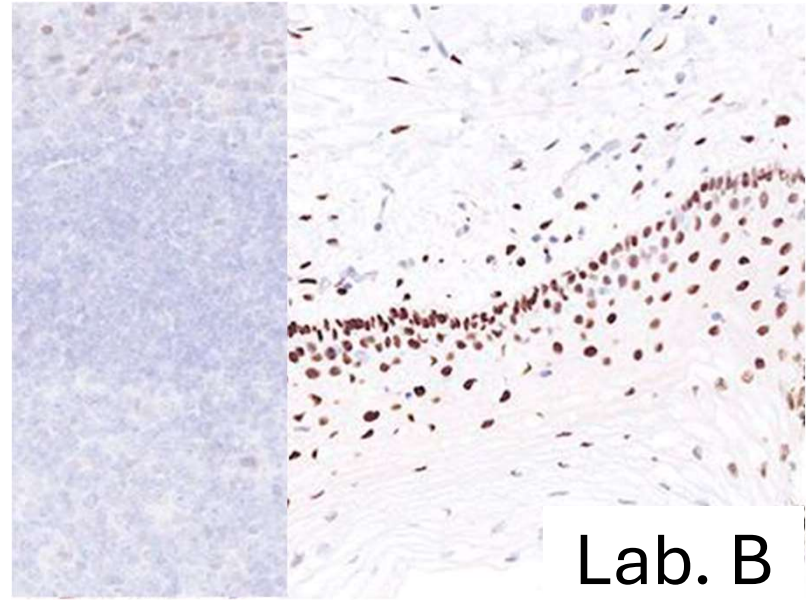
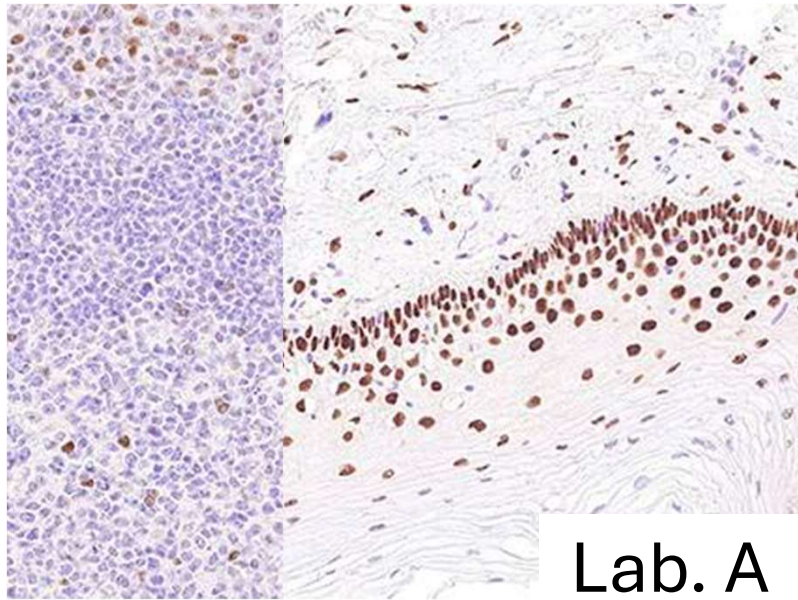
Run B33 - 2022

High  
Expressor  
Tumor



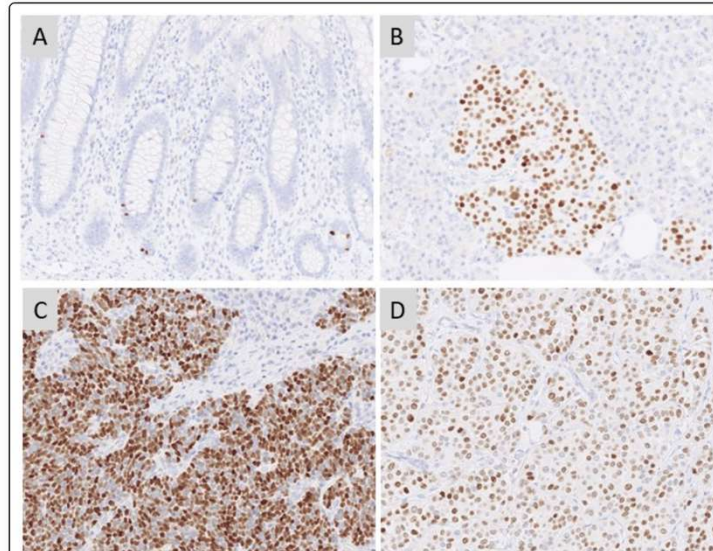
Low  
Expressor  
Tumor





# Homepage

[www.nordiqc.org](http://www.nordiqc.org)



Optimal IHC performance for INSM1:

- A. Appendix:** Neuroendocrine cells show a distinct nuclear staining reaction. Epithelial cells are negative.
- B. Pancreas:** Endocrine cells in islets of Langerhans are distinctively and selectively demonstrated.
- C. SCLC:** Virtually all neoplastic cells show a moderate to strong nuclear staining reaction.
- D. Breast NEC:** Most neoplastic cells are demonstrated indicating neuroendocrine differentiation.

## Results - Run 71 and C15

10-Jul-2024

The results for the runs 71 and C15 are now available on the website. Individual results can be seen after logging in. Protocol submission for next run is open now and deadline is 1<sup>st</sup> of September.

[All news](#)

### Events

[NordiQC Seminar in Diagnostic Immunohistochemistry 2025](#)  
17-19 Sep 2025: Aalborg, Denmark

### Important dates

[Run 72, H26, C16, B38](#)  
Publication of results  
10 Dec 2024

### Questions

Check out our [FAQ](#) (Frequently asked questions) or [contact us](#)

 [LinkedIn](#)

# NordiQC

Number of active labs: 662  
from 57 different countries.

## Participants by module

Module	n	Countries
<b>General Module</b>	509	47
<b>Breast Cancer Module</b>	519	52
<b>HER2-ISH Module</b>	293	42
<b>Companion Diagnostic Module</b>	327	40

## NordiQC assessment scheme 2023

Module	Winter	Spring	Autum
General	Run 67 <u>CD4 p40 MLH1</u> <u>CGA p53</u>	Run 68 <u>URO II/III MSH2</u> <u>TTF1 CD10</u> <u>PRAME PAX8</u>	Run 69 <u>PSA EpCAM</u> <u>CK8/18 CD5</u> <u>CD138</u>
Breast	Run B35 <u>PR HER2 IHC ER</u>		Run B36 <u>ER HER2 IHC</u>
HER2 ISH	Run H23 <u>HER2 ISH</u>		Run H24 <u>HER2 ISH</u>
Companion		Run C13 <u>PD-L1 (TPS/CPS)</u> <u>PD-L1 (IC)</u>	Run C14 <u>PD-L1 (TPS/CPS)</u> <u>PD-L1 (IC)</u>

# NordiQC method

- Construction of Tissue Micro Array block containing critical tissue for a specific epitope
- Participants submit their staining protocol on the NordiQC homepage ([www.nordiqc.org](http://www.nordiqc.org)).
- Sections are cut from the block(s) and circulated to the participants
- Participants stain the slides using the submitted protocols and return the slides to NordiQC
- A group of pathologists and expert technicians meet for assessment of the slides
- General reports describing optimal and insufficient staining protocol parameters published on [www.nordiqc.org](http://www.nordiqc.org)
- Individual assessment marks and tailored protocol recommendations sent to participants

## Modify protocol ID 635, CDX2, run 48

### Staining platform

Staining platform  ▾

### Primary antibody

Primary antibody clone  ▾

Lot number

Dilution factor : 1:400

Diluent buffer  ▾

Incubation time (minutes)

Incubation temperature (Celcius)

### Epitope Retrieval, HIER

Epitope retrieval, HIER  YES  NO

Device  ▾

HIER buffer  ▾

Efficient Heating Time (minutes)

Max. heating temperature (Celcius)

### Epitope Retrieval, proteolysis

Epitope retrieval, proteolysis  YES  NO

### Visualization system

Visualization system  ▾

Amplification  ▾

Incubation time linker (minutes)

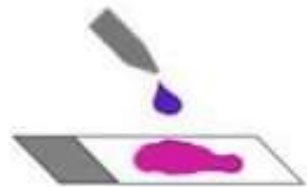
Incubation time polymer (minutes)

# NordiQC – test materials

- Multi-tissue FFPE blocks
- 10% NBF 24-48 (ASCO-CAP guidelines)
  - Normal and clinically relevant tumour tissues
  - Different levels of antigen expression (High, moderate, low, negative)

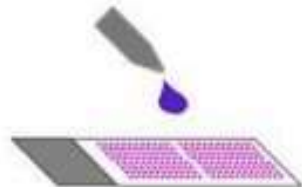


Traditional methods



1 tumor

Tissue Microarray



1000 tumors





## PReferentially expressed Antigen in Melanoma (PRAME)

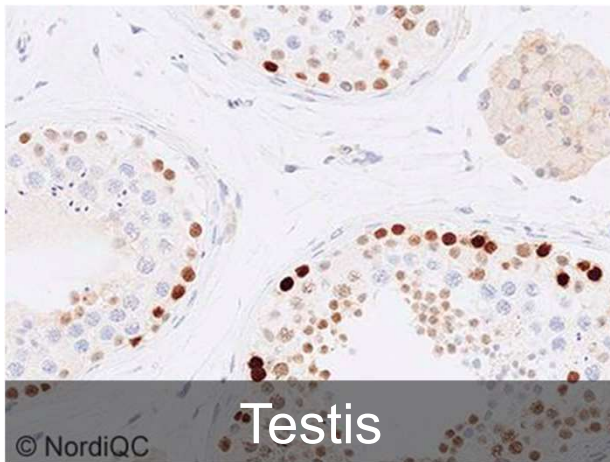
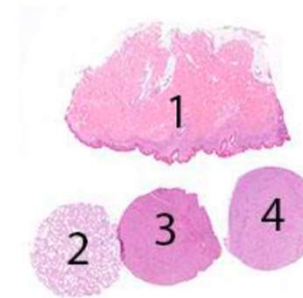
**Material**

The slide to be stained for PRAME comprised of:

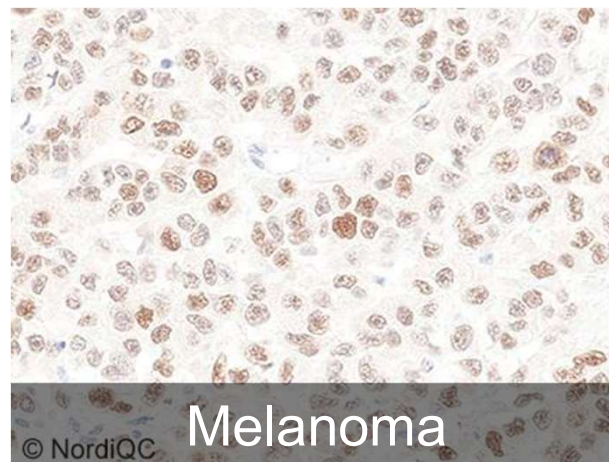
1. Compound nevus, 2. Testis, 3.-4. Melanoma.

All tissues were fixed in 10% neutral buffered formalin.

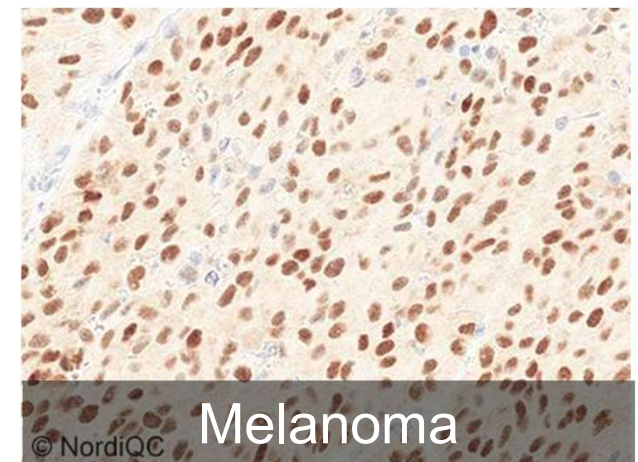
Criteria for assessing a PRAME staining as optimal included:



Testis



Melanoma



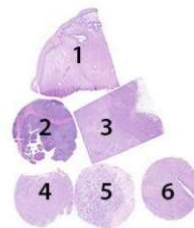
Melanoma

**Assessment Run B27 2019**  
**Estrogen receptor (ER)**

**Material**

The slide to be stained for ER comprised:

No.	Tissue	ER-positivity*	ER-intensity*
1.	Uterine cervix	80- 90%	Moderate to strong
2.	Tonsil	1-5%	Weak to moderate
3.	Breast carcinoma	70-90%	Weak to moderate
4.	Breast carcinoma	80-100%	Weak to moderate
5.	Breast carcinoma	100%	Moderate to strong
6.	Breast carcinoma	Negative	-

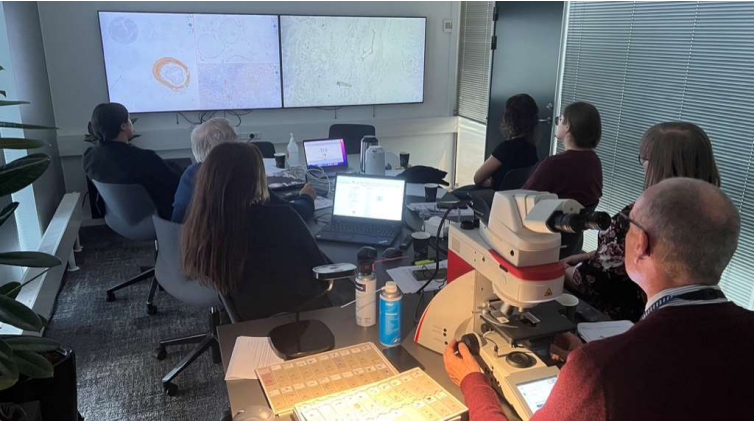


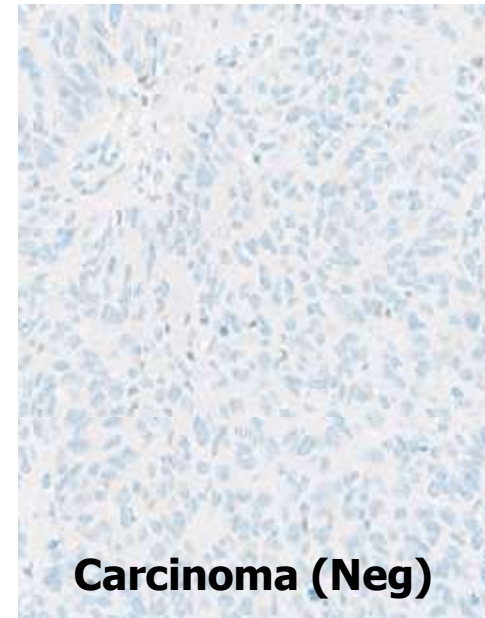
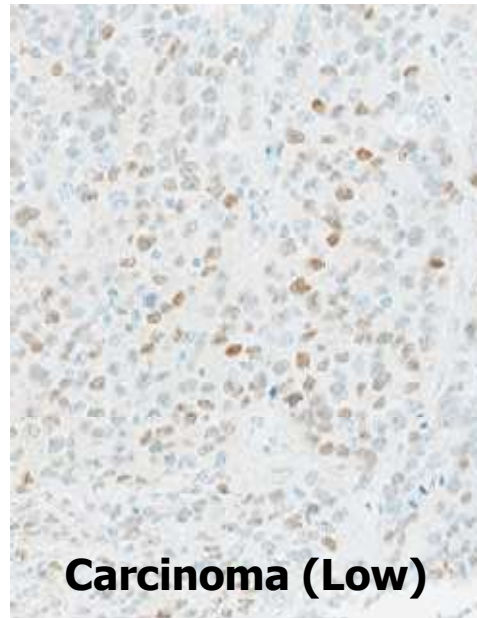
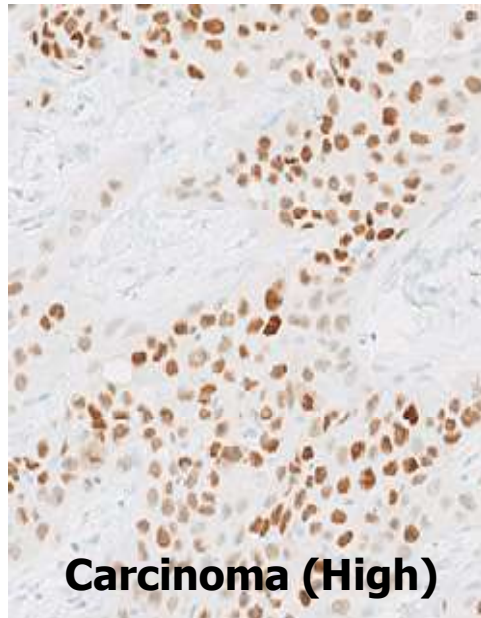
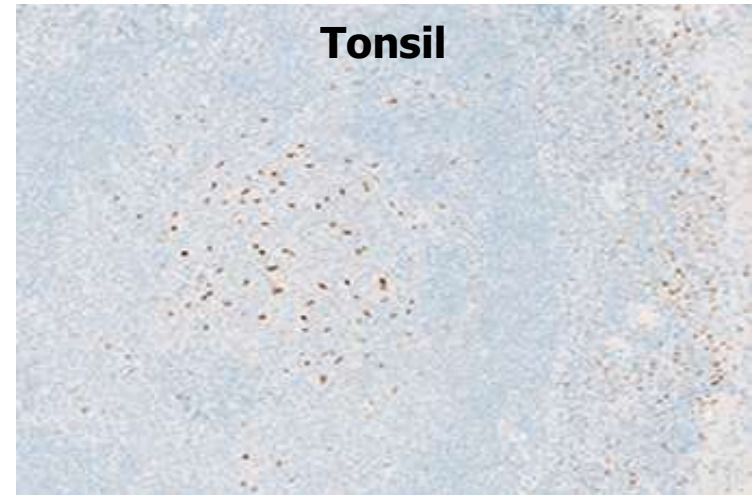
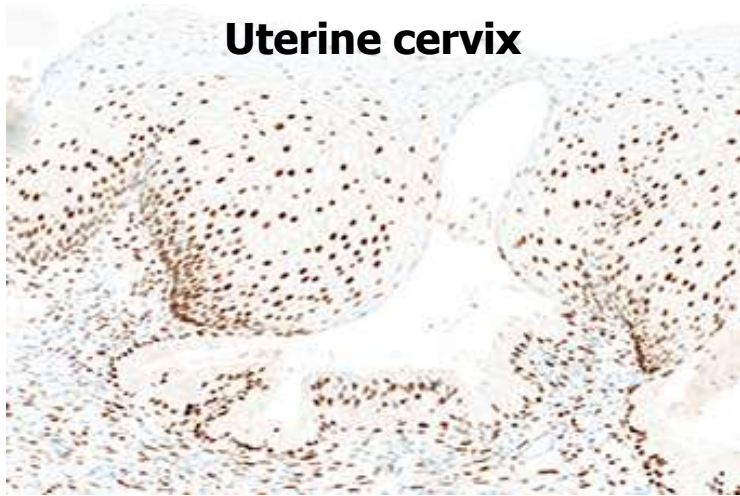
\*ER-status and staining pattern as characterized by the NordiQC reference laboratories using the rmAb clones EP1 and SP1.

Main focus of assessment:

- Appropriate technical quality (signal-to-noise, good morphology etc.)
- Appropriate analytical sensitivity and specificity – indicated by concordance of ER status and proportion of positive cells in the included tumours to references

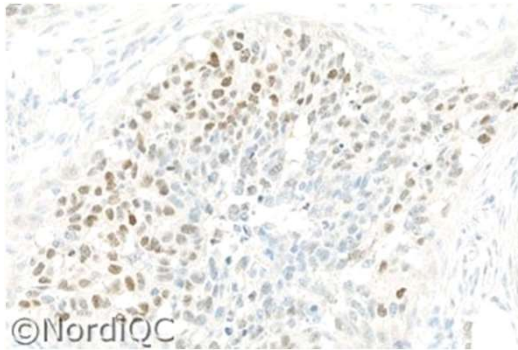
Score	Criteria: Staining reaction considered ...
Optimal	... Perfect or close to perfect in all of the included tissue cores
Good	... Fully acceptable in all of the included tissue cores. However, the protocol may be optimized to ensure the best staining intensity and signal-to-noise ratio
Borderline	... Insufficient because of, e.g., a generally too weak staining or a false negative staining of one of the included tissues, or a minor false positive staining reaction
Poor	... Very insufficient because of, e.g., false negative staining of several of the included tissues, or a major false positive staining reaction



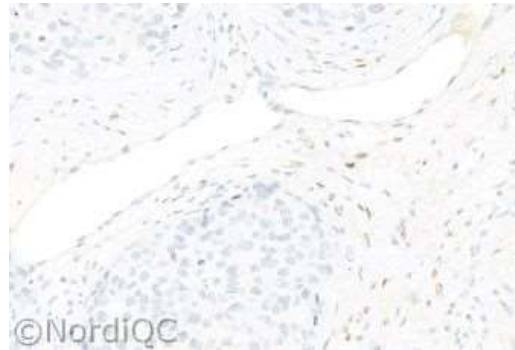


# ER: Typical challenges

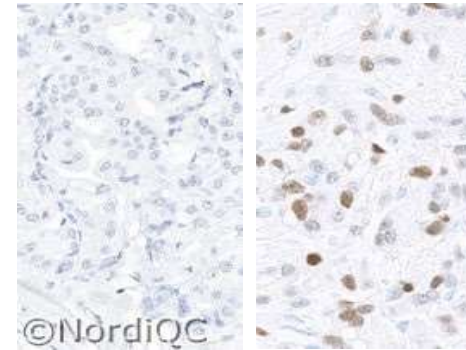
85% Weak / False negative



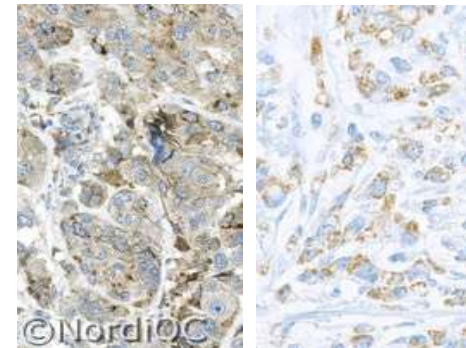
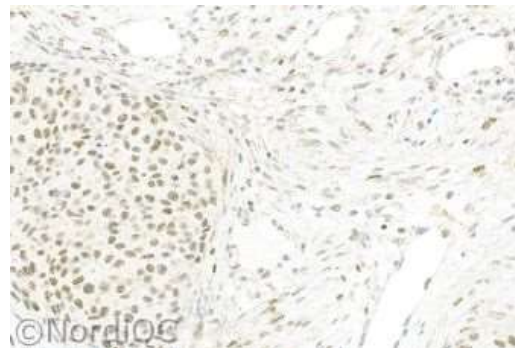
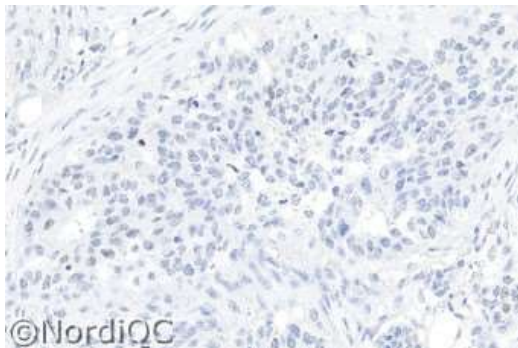
10% False positive



5% Impaired morphology, etc



**Sufficient**



**Insufficient**

Too low titre (EP1, SP1 conc.)  
Insufficient HIER,  
Clone 1D5

Clone 6F11 by HIER at high  
pH, 3-step pol.  
(not observed on VMS)

Clone 1D5 at high titre,  
Biotin-based kits,  
HIER in pressure cooker

**ER:  
Selection of  
primary Ab  
and format**

Table 1. **Antibodies and assessment marks for ER, B27**

Concentrated antibodies	n	Vendor	Optimal	Good	Borderline	Poor	Suff. <sup>1</sup>	Suff. OPS <sup>2</sup>
mAb clone <b>6F11</b>	15	Leica/Novocastra	6	6	1	2	80%	100%
mAb clone <b>C6H7</b>	1	Celnovte	-	1	-	-	-	-
rmAb clone <b>EP1</b>	16	Dako/Agilent	8	6	3	-	82%	91%
	1	Cell Marque						
	20	Thermo Scientific						
	7	Cell Marque						
rmAb clone <b>SP1</b>	1	Spring Bioscience	19	7	4	1	84%	100%
	1	Abcam						
	1	Diagnostic Biosystems						
	1	Zytomed Systems						
<b>Ready-to-Use antibodies</b>								
mAb clone <b>1D5 IR/IS657</b>	1	Dako/Agilent	1	-	-	-	-	-
mAb clones <b>1D5 + ER-2-123 SK310</b>	1	Dako/Agilent	-	1	-	-	-	-
mAb clone <b>6F11 PA0009/PA0151</b>	13	Leica	4	4	3	2	62%	83%
rmAb <b>EP1 IR/IS084</b>	27	Dako/Agilent	10	13	4	-	85%	84%
rmAb <b>EP1 IR/IS084<sup>3</sup></b>	8	Dako/Agilent	3	3	1	1	-	-
rmAb <b>EP1 GA084</b>	32	Dako/Agilent	14	15	3	-	91%	91%
rmAb <b>EP1 GA084<sup>3</sup></b>	3	Dako/Agilent	3	-	-	-	-	-
rmAb clone <b>SP1 790-4324/5</b>	187	Ventana/Roche	113	65	6	3	95%	95%
rmAb clone <b>SP1 790-4324/5<sup>2</sup></b>	1	Ventana/Roche	1	-	-	-	-	-
rmAb clone <b>SP1 249R-1</b>	4	Cell Marque	1	3	-	-	-	-
rmAb clone <b>SP1 KIT-0012</b>	1	Maixin	1	-	-	-	-	-
rmAb <b>SP1 M3011</b>	1	Spring Biosystems	-	1	-	-	-	-
rmAb clone <b>SP1 MAD-000306QD</b>	1	Master Diagnostica	-	-	1	-	-	-
rmAb clone <b>EP1 8361-C010</b>	1	Sakura Finetek	-	1	-	-	-	-
rmAb clone <b>SP1 RMPD001</b>	2	Diagnostics Biosystem	2	-	-	-	-	-
r/mAb clones <b>6F11 + SP1 PM308</b>	1	Biocare Medical	1	-	-	-	-	-
<b>Total</b>	<b>348</b>		<b>187</b>	<b>126</b>	<b>26</b>	<b>9</b>	<b>-</b>	
<b>Proportion</b>			<b>54%</b>	<b>36%</b>	<b>7%</b>	<b>3%</b>	<b>90%</b>	

1) Proportion of sufficient stains (optimal or good).  
 2) Proportion of sufficient stains with optimal protocol settings only, see below.  
 3) RTU system used on a different platform than it was developed for.

Concentrated format:  
Overall protocol parameters

HIER alk. pH  
2- & 3-step kits

Carefully calibration of primary Ab

# Causes of insufficient results

- Less successful antibodies
  - Poor antibodies
  - Less robust antibodies
  - Poorly calibrated RTUs
  - Stainer platform dependent antibodies
- Insufficiently calibrated antibody dilutions
- Insufficient or erroneous epitope retrieval
- Less sensitive visualization systems
- Other
  - Impaired morphology
  - Technical issues
  - Excessive counterstaining impairing interpretation

# Individual feedback



## Nordic Immunohistochemical Quality Control

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### Assessment of run 54 - individual results

Klinik XXX

Epitope	Podop	GATA3	CK-PAN	CEA
Assessment	Poor	Poor	Good	Good

#### Comments

##### **Podop - Poor**

Comment: False negative.

Advice: Increase primary Ab conc. Consider change of HIER buffer to Tris-EDTA pH 9 or equivalent - cave biotin.

##### **GATA3 - Poor**

Comment: False negative. False positive.

Advice: Use Hier in alkaline buffer and/or change to a more sensitive detection system and recalibrate portocol settings .

##### **CK-PAN - Good**

Comment: Weak.

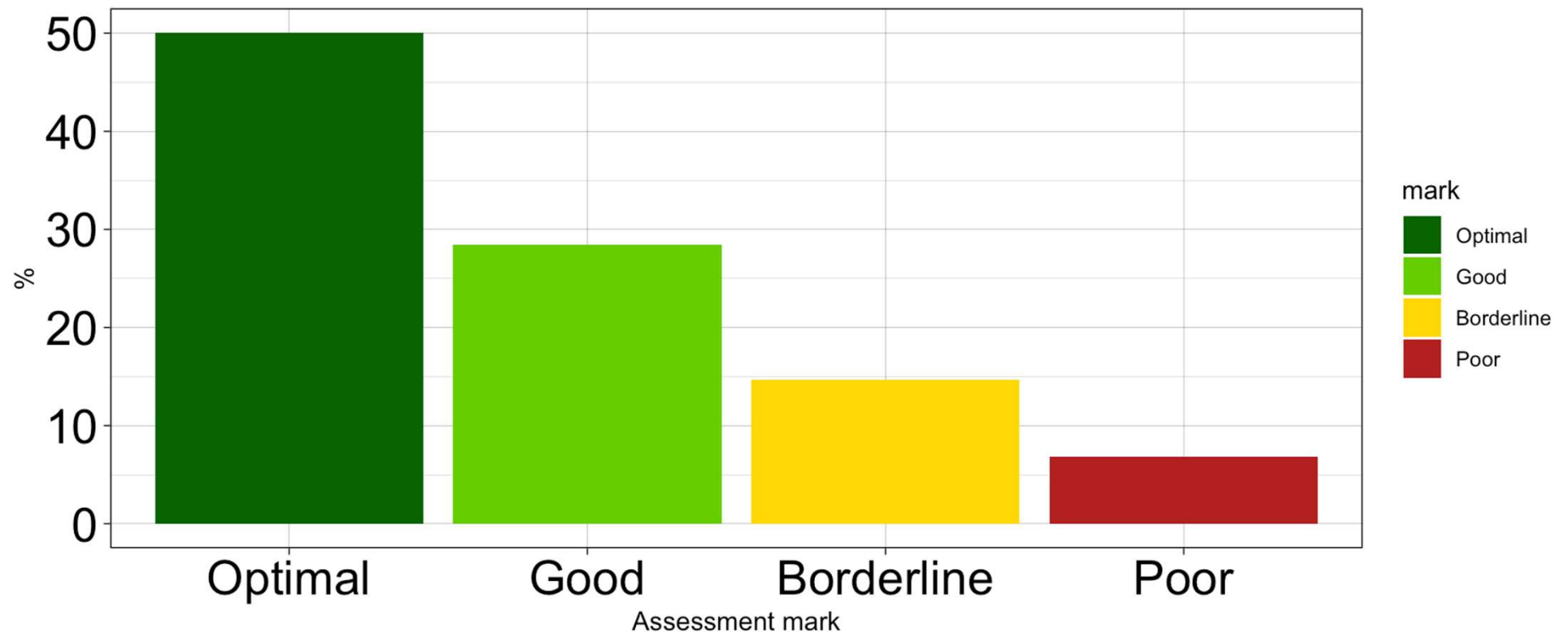
##### **CEA - Good**

Comment: Weak. Excessive background.

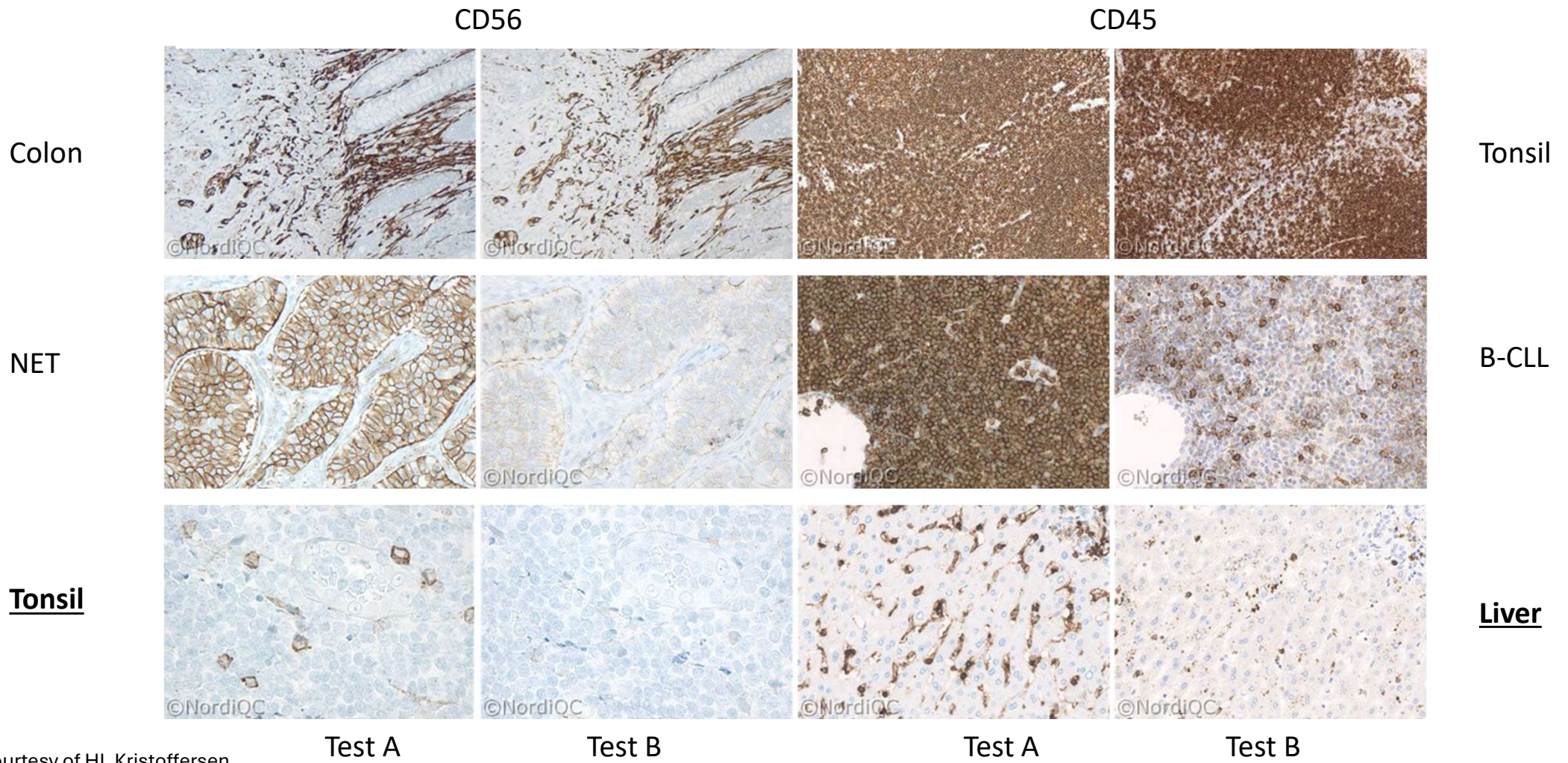




# Results – assessment marks 2016-24



# Selection of controls is imperative for IHC quality


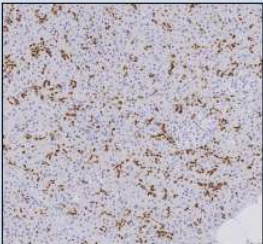



# NordiQC IHC control atlas



Info ▾ Modules ▾ Assessments Protocols Controls Events ▾ [SN](#)

## CDX2 - CDX2

Control type	Positive tissue control High expression level	Positive tissue control Low expression levels	Negative tissue control
Tissue	Appendix/colon	Pancreas	Tonsil
Description	<p>All epithelial cells must show a strong nuclear staining reaction.</p> <p><i>Note, a weak cytoplasmic staining reaction in CDX2 positive cells can be seen and should be accepted if signal-to-noise ratio otherwise is acceptable.</i></p>	<p>The vast majority of epithelial cells of intercalated ducts must show a weak to moderate nuclear staining reaction.</p>	<p>No staining reaction should be seen.</p> <p><i>Note, dispersed lymphocytes can show a faint nuclear staining reaction.</i></p>
Example	 <p>Click to enlarge</p>	 <p>Click to enlarge</p>	 <p>Click to enlarge</p>

Available for NordiQC participants

Tissues

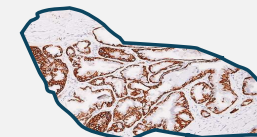
Purpose

Reaction patterns

Online scans accessible

11145  
IHC

T  
P  
A



# Conclusions

- EQA
  - Provides objective evidence of lab performance
  - Helps identify methodological errors
- Around 20-25% of slides submitted to NordiQC are still insufficient!
- Labs not participating in EQA ?
- How many scientific publications are based on insufficient IHC stains?
- What are the consequences for patients?

# Thank you for your attention!

## Collaborators in NordiQC

Søren Nielsen  
Heidi Lykke Kristoffersen  
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Louise Rønnow Holler  
Michael Bzorek  
Tanya Julio  
Kristi Bøgh Andersson

And the whole assessor team!

